

Docket No. 203979US0

Title of the Invention

NUCLEOTIDE SEQUENCES CODING FOR THE *lysR1* GENE

Cross-Reference to Related Application

- 5 The present application claims priority to German Application No. DE 100 39 044.7 filed August 10, 2000, the entire contents of which are incorporated herein by reference.

BACKGROUND OF THE INVENTION

Field of the Invention

The invention provides nucleotide sequences from *Coryneform* bacteria which code for the *lysR1* gene and a process for the fermentative preparation of amino acids, in particular L-lysine by attenuation of the *lysR1* gene. The *lysR1* gene codes for the LysR1 protein, which is a transcription regulator of the LysR family.

Discussion of the Background

- 20 L-amino acids, in particular L-lysine, are used in human medicine and in the pharmaceutical industry, in the foodstuffs industry, and most particularly in animal nutrition.

- 25 It is known that amino acids can be produced by fermentation of strains of *Coryneform* bacteria, in particular *Corynebacterium glutamicum*. On account of the great importance of these amino acids constant efforts are being made to improve the production processes. Improvements in production processes may involve fermentation technology measures, such as for example stirring and provision of oxygen, or the composition of the

5 nutrient media, such as for example the sugar concentration during the fermentation, or the working up to the product form by for example ion exchange chromatography, or the intrinsic performance properties of the microorganism itself.

10 Methods involving mutagenesis, selection and mutant selection are used to improve the performance properties of these microorganisms. In this way strains are obtained that are resistant to antimetabolites or are auxotrophic for regulatorily significant metabolites and that produce amino acids.

15 Methods of recombinant DNA have also been employed for improving strains of *Corynebacterium* strains which produce L-amino acids.

20 However, there remains a critical need for improved methods of producing L-amino acids and thus for the provision of strains of bacteria producing higher amounts of L-amino acids. On a commercial or industrial scale even small improvements in the yield of L-amino acids, or the efficiency of their production, are economically significant. Prior to the present invention, it was not recognized that attenuation of lysR1 gene encoding the a LysR1 transcriptional regulation protein would improve L-amino acid yields.

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SUMMARY OF THE INVENTION

30 One object of the present invention, is providing a new process adjuvant for improving the fermentative production of L-amino acids, particularly L-lysine and L-glutamate. Such process adjuvants include enhanced bacteria, preferably enhanced *Coryneform* bacteria which express attenuated amounts of LysR1 transcriptional regulator which is encoded by the lysR1 gene.

Thus, another object of the present invention is providing such an bacterium, which expresses an attenuated amount of LysR1 transcriptional regulator or gene products of the lysR1 gene.

- 5 Another object of the present invention is providing a bacterium, preferably a *Coryneform* bacterium, which expresses a polypeptide that has an attenuated LysR1 transcriptional regulator activity.

10 Another object of the invention is to provide a nucleotide sequence encoding a polypeptide which has LysR1 transcriptional regulator sequence. One embodiment of such a sequence is the nucleotide sequence of SEQ ID NO: 1.

15 A further object of the invention is a method of making LysR3 transcriptional regulator or an isolated polypeptide having a LysR1 transcriptional regulator activity, as well as use of such isolated polypeptides in the production of amino acids. One embodiment of such a polypeptide is the polypeptide having the amino acid sequence of SEQ ID NO: 2.

20 Other objects of the invention include methods of detecting nucleic acid sequences homologous to SEQ ID NO: 1, particularly nucleic acid sequences encoding polypeptides that have LysR1 transcriptional regulator activity, and methods of making nucleic acids encoding such polypeptides.

25 The above objects highlight certain aspects of the invention. Additional objects, aspects and embodiments of the invention are found in the following detailed description of the invention.

BRIEF DESCRIPTION OF THE DRAWING

Figure 1: Map of the plasmid pCR2.1lysR1int.

DETAILED DESCRIPTION OF THE INVENTION

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art of molecular biology. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described herein. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In addition, the materials, methods, and examples are illustrative only and are not intended to be limiting.

Reference is made to standard textbooks of molecular biology that contain definitions and methods and means for carrying out basic techniques, encompassed by the present invention. See, for example, Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York (1982) and Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York (1989) and the various references cited therein.

As used herein, L-amino acids or amino acids are understood to mean and amino acid or its salt. Preferably, the amino acids are chosen from the group consisting of L-asparagine, L-threonine, L-serine, L-glutamate, L-glycine, L-alanine, L-cysteine, L-valine, L-methionine, L-isoleucine, L-leucine, L-tyrosine, L-phenylalanine, L-histidine, L-lysine, L-tryptophan and L-arginine. L-Lysine is particularly preferred.

As used herein L-lysine or lysine include not only the bases but also the salts, such as e.g. lysine monohydrochloride or lysine sulfate.

The invention provides an isolated polynucleotide of
Coryneform bacteria containing a polynucleotide sequence
5 coding for the *lysR1* gene, selected from the group
comprising

- a) polynucleotide that is at least 70% identical to a
polynucleotide coding for a polypeptide that contains
the amino acid sequence of SEQ ID No. 2,
- b) polynucleotide coding for a polypeptide that contains an
amino acid sequence that is at least 70% identical to
the amino acid sequence of SEQ ID No. 2,
- c) polynucleotide that is complementary to the
polynucleotides of a) or b), and
- d) polynucleotide containing at least 15 successive
nucleotides of the polynucleotide sequence of a), b) or
c),

the polypeptide preferably having the activity of the
transcription regulator *lysR1*.

20 The invention also provides the aforementioned
polynucleotide, which is preferably a replicable DNA
containing:

- (i) the nucleotide sequence shown in SEQ ID No.1, or
- (ii) at least one sequence that corresponds to the
25 sequence (i) within the region of degeneration of
the genetic code, or
- (iii) at least one sequence that hybridises with the
sequences that are complementary to the sequences
(i) or (ii), and optionally

(iv) functionally neutral sense mutations in (i).

The invention furthermore provides:

a replicable DNA containing the nucleotide sequence as illustrated in SEQ ID No.1;

5 a polynucleotide coding for a polypeptide that contains the amino acid sequence as is illustrated in SEQ ID No. 2;

a vector containing the polynucleotide d) according to the invention, in particular pCR2.llysRlint inserted into *E. Coli* DSM 13616 and filed at DSMZ, Brunswick, (Germany);

and *Coryneform* bacteria that in the lysR1 gene contain an insertion or deletion, in particular by using the vector pCR2.llysRlint.

The invention thus provides polynucleotides consisting substantially of a polynucleotide sequence, that are obtainable by screening by hybridising a corresponding gene library that contains the complete gene with the polynucleotide sequence corresponding to SEQ ID No.1, with a probe that contains the sequence of the aforementioned polynucleotide according to SEQ ID No. 1 or a fragment thereof, and isolating the aforementioned DNA sequence.

Polynucleotide sequences according to the invention are suitable as hybridisation probes for RNA, cDNA and DNA, in order to isolate nucleic acids, polynucleotides or genes in their full length that code for lysR1 protein, or to isolate such nucleic acids or polynucleotides or genes that have a high degree of similarity to the sequence of the lysR1 gene.

Polynucleotide sequences according to the invention are further suitable as primers with the polymerase chain

reaction (PCR), DNA of genes can be produced that code for lysR1 protein.

Such oligonucleotides serving as probes or primers contain at least 30, preferably at least 20, and most particularly preferably at least 15 successive nucleotides. Also suitable are oligonucleotides having a length of at least 40 or 50 nucleotides.

"Isolated" denotes separated from its natural environment.

10 "Polynucleotide" refers in general to polyribonucleotides and polydeoxyribonucleotides, which may either be unmodified RNA or DNA or modified RNA or DNA.

The term "polypeptides" denotes peptides or proteins that contain two or more amino acids bound via peptide bonds.

15 The polypeptides according to the invention include a polypeptide according to SEQ ID No. 2, in particular those peptides having the biological activity of lysR1 protein, and also those polypeptides that are at least 70%, preferably at least 80% and particularly preferably at least 90% to 95% identical to the polypeptide according to
20 SEQ ID No. 2 and have the aforementioned activity.

The present invention furthermore relates to a process for the enzymatic production of amino acids, in particular L-lysine, using *Coryneform* bacteria that in particular already produce amino acids and in which the nucleotide
25 sequences coding for the lysR1 gene are attenuated, in particular are switched off or are expressed at a low level.

30 The term "attenuation" used in this context denotes the reduction or switching off of the intracellular activity of one or more enzymes (proteins) in a microorganism that are coded by the corresponding DNA, by for example using a weak promoter or using a gene or allele that codes for a

corresponding gene having a low activity or that inactivates the corresponding gene or enzyme (protein), and optionally combining these measures.

5 The microorganisms that are the subject of the present invention may produce amino acids, in particular L-lysine, from glucose, sucrose, lactose, fructose, maltose, molasses, starch, cellulose or from glycerol and ethanol. These microorganisms may be representatives of *Coryneform* bacteria, in particular of the genus *Corynebacterium*. In
10 the genus *Corynebacterium* the species *Corynebacterium glutamicum* should in particular be mentioned, which is known to those skilled in the art for its ability to produce L-amino acids.

Suitable strains of the genus *Corynebacterium*, in particular of the species *Corynebacterium glutamicum* (*C. glutamicum*), are in particular the known wild type strains

Corynebacterium glutamicum ATCC13032
Corynebacterium acetoglutamicum ATCC15806
Corynebacterium acetoacidophilum ATCC13870
20 *Corynebacterium melassecola* ATCC17965
Corynebacterium thermoaminogenes FERM BP-1539
Brevibacterium flavum ATCC14067
Brevibacterium lactofermentum ATCC13869 and
Brevibacterium divaricatum ATCC14020

25 or mutants or strains formed therefrom that produce L-amino acids, such as for example the strains producing L-lysine.

Corynebacterium glutamicum FERM-P 1709
Brevibacterium flavum FERM-P 1708
Brevibacterium lactofermentum FERM-P 1712
30 *Corynebacterium glutamicum* FERM-P 6463
Corynebacterium glutamicum FERM-P 6464
Corynebacterium glutamicum DM58-1
Corynebacterium glutamicum DG52-5

Corynebacterium glutamicum DSM 5714 and
Corynebacterium glutamicum DSM 12866

Preferably, a bacterial strain with attenuated expression
5 of a *lysR1* gene that encodes a polypeptide with *LysR1*
transcriptional regulation activity will improve amino acid
yield at least 1%.

The inventors have successfully isolated the new *lysR1* gene
from *C. glutamicum* coding for *lysR1* protein, which is a
transcription regulator of the *lysR* family.

In order to isolate the *lysR1* gene or also other genes from
C. glutamicum, a gene library of this microorganism is
first of all introduced into *Escherichia coli* (*E. coli*).
The introduction of gene libraries is described in
generally known textbooks and manuals. As an example there
15 may be mentioned the textbook by Winnacker: *Gene and Klone*,
Eine Einführung in die Gentechnologie (Verlag Chemie,
Weinheim, Germany, 1990), or the manual by Sambrook et al.:
Molecular Cloning, A Laboratory Manual (Cold Spring Harbor
Laboratory Press, 1989). A very well-known gene library is
20 that of the *E. coli* K-12 strain W3110, which was introduced
by Kohara et al. (*Cell* 50, 495-508 (1987)) into λ -vectors.
Bathe et al. (*Molecular and General Genetics*, 252:255-265,
1996) describe a gene library of *C. glutamicum* ATCC13032,
25 which was introduced by means of the cosmid vector SuperCos
I (Wahl et al., 1987, *Proceedings of the National Academy*
of Sciences USA, 84:2160-2164) in the *E. coli* K-12 strain
NM554 (Raleigh et al., 1988, *Nucleic Acids Research*
16:1563-1575). Börmann et al. (*Molecular Microbiology*
30 6(3), 317-326 (1992)) again describe a gene library of *C.*
glutamicum ATCC13032 using the cosmid pH79 (Hohn and
Collins, 1980, *Gene* 11, 291-298).

In order to produce a gene library of *C. glutamicum* in *E. coli* plasmids such as pBR322 (Bolivar, 1979, Life Sciences, 25, 807-818) or pUC9 (Vieira et al., 1982, Gene, 19:259-268) may also be employed. Suitable hosts are in

5 particular those *E. coli* strains that are restriction and recombinant defective, for example the strain DH5 α , (Jeffrey H. Miller: "A Short Course in Bacterial Genetics, A Laboratory Manual and Handbook for *Escherichia coli* and Related Bacteria", Cold Spring Harbor Laboratory Press, 10 1992)

The long DNA fragments cloned with the help of cosmids or other λ -vectors may then in turn be subcloned into conventional vectors suitable for DNA sequencing.

Methods of DNA sequencing are described in, *inter alia*, Sanger et al. (Proceedings of the National Academy of Sciences of the United States of America USA, 74:5463-5467, 1977).

The DNA sequences obtained may then be investigated with known algorithms or sequence analysis programs, such as for example that of Staden (Nucleic Acids Research 14, 217-232(1986)), that of Marck (Nucleic Acids Research 16, 1829-1836 (1988)) or the GCG program of Butler (Methods of Biochemical Analysis 39, 74-97 (1998)).

25 The new DNA sequence of *C. glutamicum* coding for the lysR1 gene was obtained in this way, and as SEQ ID No. 1 is part of the present invention. The amino acid sequence of the corresponding protein was also derived from the existing DNA sequence using the aforescribed methods. The resulting amino acid sequence of the lysR1 gene product is 30 shown in SEQ ID No. 2.

Coding DNA sequences that are obtained from SEQ ID No. 1 as a result of the degenerability of the genetic code are also covered by the invention. Similarly, DNA sequences that

hybridise with SEQ ID No. 1 or parts of SEQ ID No. 1 are also covered by the invention. Furthermore, in this specialist field conservative amino acid replacements, such as for example the replacement of glycine by alanine or of aspartic acid by glutamic acid in proteins, are known as sense mutations, which do not lead to any fundamental change in the activity of the protein, i.e. are functionally neutral. Furthermore, it is known that changes at the N-terminus and/or C-terminus of a protein do not significantly impair or may even stabilise its function. Those skilled in the art can find details of this in, *inter alia*, Ben-Bassat et al. (Journal of Bacteriology 169:751-757 (1987)), in O'Regan et al. (Gene 77:237-251 (1989)), in Sahin-Toth et al. (Protein Sciences 3:240-247 (1994)), in Hochuli et al. (Bio/Technology 6:1321-1325 (1988)) and in known textbooks of genetics and molecular biology. Amino acid sequences that are obtained in a corresponding manner from SEQ ID No. 2 are likewise covered by the invention.

Finally, DNA sequences that are produced by the polymerase chain reaction (PCR) using primers resulting from SEQ ID No. 1, are also covered by the invention. Such oligonucleotides typically have a length of at least 15 nucleotides.

The person skilled in the art can find details of the identification of DNA sequences by means of hybridisation in, *inter alia*, the textbook "The DIG System User's Guide for Filter Hybridization" published by Boehringer Mannheim GmbH (Mannheim, Germany, 1993) and in Liebl et al.

(International Journal of Systematic Bacteriology 41: 255-260 (1991)). The person skilled in the art can obtain details of the amplification of DNA sequences by means of the polymerase chain reaction (PCR) in, *inter alia*, the handbook by Gait: Oligonucleotide Synthesis: A Practical Approach (IRL Press, Oxford, UK, 1984) and the Newton and

Graham: PCR (Spektrum Akademischer Verlag, Heidelberg, Germany, 1994).

In the course of work carried out on the present invention it was found that *Coryneform* bacteria after attenuation of the lysR1 gene produce amino acids, in particular L-lysine, in an improved manner.

In order to achieve an attenuation, either the expression of the lysR1 gene or the catalytic properties of the enzyme protein may be reduced or switched off. Optionally both measures may be combined.

The reduction of the gene expression may be achieved by suitable culture conditions or by genetic alteration (mutation) of the signal structures of the gene expression. Signal structures of the gene expression are for example repressor genes, activator genes, operators, promoters, attenuators, ribosome binding sites, the start codon and terminators. The person skilled in the art can obtain further information on this in for example patent application WO 96/15246, in Boyd and Murphy (Journal of Bacteriology 170: 5949 (1988)), in Voskuil and Chambliss (Nucleic Acids Research 26: 3548 (1998)), in Jensen and Hammer (Biotechnology and Bioengineering 58: 191 (1998)), in Pátek et al. (Microbiology 142: 1297 (1996)), Vasicova et al. (Journal of Bacteriology 181: 6188 (1999)) and in known textbooks of genetics and molecular biology, such as for example the textbook by Knippers ("Molekulare Genetik", 6th Edition, Georg Thieme Verlag, Stuttgart, Germany, 1995) or the textbook by Winnacker ("Gene and Klone", VCH Verlagsgesellschaft, Weinheim, Germany, 1990).

Mutations that lead to an alteration or reduction of the catalytic properties of enzyme proteins are known in the prior art; as examples there may be mentioned the work of Qiu and Goodman (Journal of Biological Chemistry 272: 8611-8617 (1997)), Sugimoto et al. (Bioscience Biotechnology and

Biochemistry 61: 1760-1762 (1997)) and Möckel ("Die Threonindehydratase aus *Corynebacterium glutamicum*: Aufhebung der allosterischen Regulation and Struktur des Enzyms", and reports published by the Jülich Research Centre, Jül-2906, ISSN09442952, Jülich, Germany, 1994). Overviews may be obtained from known textbooks on genetics and molecular biology, for example that of Hagemann ("Allgemeine Genetik", Gustav Fischer Verlag, Stuttgart, 1986).

Mutations in the present context include transitions, transversions, insertions and deletions. Depending on the effect of the amino acid replacement on the enzyme activity, one talks either of missense mutations or nonsense mutations. Insertions or deletions of at least one base pair (bp) in a gene lead to frame shift mutations, following which false amino acids are incorporated or the translation terminates prematurely. Deletions of several codons typically lead to a complete cessation of enzyme activity. Details of the production of such mutations are part of the prior art and may be obtained from known textbooks on genetics and molecular biology, such as for example the textbook by Knippers ("Molekulare Genetik", 6th Edition, Georg Thieme Verlag, Stuttgart, Germany, 1995), the textbook by Winnacker ("Gene and Klone", VCH Verlagsgesellschaft, Weinheim, Germany, 1990) or the textbook by Hagemann ("Allgemeine Genetik", Gustav Fischer Verlag, Stuttgart, 1986).

A conventional method of mutating genes of *C. glutamicum* is the method of gene disruption and gene replacement described by Schwarzer and Pühler (Bio/Technology 9, 84-87 (1991)).

In the method of gene disruption a central part of the coding region of the gene in question is cloned into a plasmid vector that can replicate in a host (typically *E. coli*), but not in *C. glutamicum*. Suitable vectors are for

example pSUP301 (Simon et al., Bio/Technology 1, 784-791 (1983)), pK18mob or pK19mob (Schäfer et al., Gene 145, 69-73 (1994)), pK18mobsacB or pK19mobsacB (Jäger et al., Journal of Bacteriology 174: 5462-65 (1992)), pGEM-T (Promega Corporation, Madison, WI, USA), pCR2.1-TOPO (Shuman (1994), Journal of Biological Chemistry 269:32678-84; US-Patent 5,487,993), pCR®Blunt (Invitrogen, Groningen, Netherlands; Bernard et al., Journal of Molecular Biology, 234: 534-541 (1993)) or pEM1 (Schrumpf et al, 1991, Journal of Bacteriology 173:4510-4516). The plasmid vector that contains the central part of the coding region of the gene is then converted by conjugation or transformation into the desired strain of *C. glutamicum*. The method of conjugation is described for example by Schäfer et al. (Applied and Environmental Microbiology 60, 756-759 (1994)). Methods of transformation are described for example in Thierbach et al. (Applied Microbiology and Biotechnology 29, 356-362 (1988)), Dunican and Shivnan (Bio/Technology 7, 1067-1070 (1989)) and Tauch et al. (FEMS Microbiological Letters 123, 343-347 (1994)). After homologous recombination by means of a cross-over event, the coding region of the relevant gene is disrupted by the vector sequence and two incomplete alleles are obtained, missing respectively the 3'- and 5'-end. This method has been used for example by Fitzpatrick et al. (Applied Microbiology and Biotechnology 42, 575-580 (1994)) to switch off the *recA* gene of *C. glutamicum*.

Fig. 1 shows for example the plasmid vector pCR2.1 *lysR*int, by means of which the *lysR* gene can be disrupted or switched off.

In the gene replacement method a mutation, such as for example a deletion, insertion or base replacement, is produced *in vitro* in the gene that is of interest. The resultant allele is in turn cloned into a non-replicative vector for *C. glutamicum*, and this is then converted by

transformation or conjugation into the desired host of *C. glutamicum*. After homologous recombination by means of a first cross-over event effecting integration, and an appropriate second cross-over event effecting an excision, the incorporation of the mutation or allele in the target gene or in the target sequence is achieved. This method has been used for example by Peters-Wendisch et al. (Microbiology 144, 915 - 927 (1998)) to switch off the *pyc* gene of *C. glutamicum* by a deletion.

A deletion, insertion or a base replacement can be incorporated into the *lysR1* gene in this way.

In addition, it may be advantageous for the production of L-amino acids, in particular L-lysine, in addition to the attenuation of the *lysR1* gene, also to enhance, in particular overexpress, one or more enzymes of the respective biosynthesis pathway, glycolysis, anapleurosis, pentose phosphate cycle, or amino acid export.

Thus for example, for the production of L-lysine one or more of the genes selected from the following group may simultaneously be enhanced, in particular overexpressed

- the gene *dapA* coding for dihydrodipicolinate synthase (EP-B 0 197 335),
- the gene *eno* coding for enolase (DE: 19947791.4),
- the gene *zwf* coding for the *zwf* gene product (JP-A-09224661),
- the gene *pyc* coding for pyruvate carboxylase (Peters-Wendisch et al. (Microbiology 144, 915 - 927 (1998)))
- the gene *lysE* coding for lysine export (DE-A-195 48 222).

Also, it may be advantageous for the production of amino acids, especially L-lysine, besides attenuating the lysR1 gene, at the same time to attenuate one or more of the genes selected from the group

- 5 • the gene pck coding for phosphoenol pyruvate carboxykinase (DE 199 50 409.1, DSM 13047),
- the gene pgi coding for glucose-6-phosphate isomerase (US 09/396,478, DSM 12969),
- the gene poxB coding for pyruvate oxidase (DE:1995 1975.7, DSM 13114)

Moreover, it may be advantageous for the production of amino acids, in particular L-lysine, in addition to attenuating the lysR1 gene also to switch off undesirable secondary reactions (Nakayama: "Breeding of Amino Acid Producing Microorganisms", in: Overproduction of Microbial Products, Krumphanzl, Sikyta, Vanek (eds.), Academic Press, London, UK, 1982).

The microorganisms produced according to the invention are likewise covered by the invention and for the purposes of producing L-amino acids, in particular L-lysine, may be cultivated continuously or batchwise in a batch process, or in a feed batch process or repeated batch process. A summary of known cultivation methods is described in the textbook by Chmiel (Bioprozesstechnik 1. Einführung in die Bioverfahrenstechnik (Gustav Fischer Verlag, Stuttgart, 1991)) or in the textbook by Storhas (Bioreaktoren und periphere Einrichtungen (Vieweg Verlag, Brunswick/Wiesbaden, 1994)).

The culture medium to be used must satisfy in an appropriate manner the requirements of the respective strains. Descriptions of culture media for various microorganisms are given in the handbook "Manual of Methods for General Bacteriology" of the American Society for

Bacteriology (Washington D.C., USA, 1981). As carbon source there may be used sugars and carbohydrates such as for example glucose, sucrose, lactose, fructose, maltose, molasses, starch and cellulose, oils and fats such as for example soya oil, sunflower oil, groundnut oil and coconut oil, fatty acids such as for example palmitic acid, stearic acid and linoleic acid, alcohols such as for example glycerol and ethanol, and organic acids such as for example acetic acid. These substances may be used individually or as a mixture.

As nitrogen source there may be used organic nitrogen-containing compounds such as peptones, yeast extract, meat extract, malt extract, corn steep liquor, soya bean flour and urea, or inorganic compounds such as ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate and ammonium nitrate. The nitrogen sources may be used individually or as a mixture.

As phosphorus source there may be used phosphoric acid, potassium dihydrogen phosphate or dipotassium hydrogen phosphate, or the corresponding sodium-containing salts. The culture medium must furthermore contain salts of metals, such as for example magnesium sulfate or iron sulfate, that are necessary for growth. Finally, essential growth promoters such as amino acids and vitamins may in addition be added to the aforementioned substances. Suitable precursors may moreover be added to the culture medium. The aforementioned starting substances may be added to the culture in the form of a single batch, or metered in in an appropriate manner during the cultivation procedure.

In order to control the pH of the culture basic compounds such as sodium hydroxide, potassium hydroxide, ammonia or ammonia water, or acidic compounds such as phosphoric acid or sulfuric acid may be added in an appropriate manner. In order to control foam formation anti-foaming agents such as

for example fatty acid polyglycol esters may be used. In order to maintain the stability of plasmids selectively acting substances, such as for example antibiotics, may be added to the medium. In order to maintain aerobic
5 conditions, oxygen or oxygen-containing gas mixtures, such as for example air, are pumped into the culture. The temperature of the culture is normally 20°C to 45°C and preferably 25°C to 40°C. The cultivation is continued until a maximum amount of the desired product has been
10 formed. This target is normally reached within 10 hours to 160 hours.

Methods for determining L-amino acids are known from the prior art. The analysis may for example be carried out as described by Spackman et al. (Analytical Chemistry, 30, (1958), 1190) by anion exchange chromatography followed by ninhydrin derivatisation, or it may be carried out by reversed phase HPLC, as described by Lindroth et al. (Analytical Chemistry (1979) 51: 1167-1174).

The following microorganism has been filed according to the Budapest Convention at the German Collection for Microorganisms and Cell Cultures (DSMZ, Brunswick, Germany).

- *Escherichia coli* strain *E. coli* TOP10F/pCR2.11lysR1int as DSM 13616.

25 The process according to the invention serves for the enzymatic production of amino acids, in particular L-lysine.

The present invention is illustrated in more detail hereinafter with the aid of examples of implementation.

30 The isolation of plasmid DNA from *Escherichia coli* as well as all techniques for the restriction, Klenow and alkaline phosphatase treatment were carried out according to Sambrook et al. (Molecular Cloning. A Laboratory Manual,

1989, Cold Spring Harbour Laboratory Press, Cold Spring Harbor, NY, USA). Methods for the transformation of *Escherichia coli* are likewise described in this handbook.

5 The compositions of conventional nutrient media such as LB medium or TY medium may also be obtained from the handbook by Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York (1989).

Example 1

10 Production of a genomic cosmid gene library from *C. glutamicum* ATCC 13032

15 Chromosomal DNA from *C. glutamicum* ATCC 13032 was isolated as described by Tauch et al., (1995, Plasmid 33:168-179) and partially cleaved with the restriction enzyme Sau3AI (Amersham Pharmacia, Freiburg, Germany, Product Description Sau3AI, Code no. 27-0913-02). The DNA fragments were dephosphorylated with shrimp alkaline phosphatase (Roche Molecular Biochemicals, Mannheim, Germany, Product Description SAP, Code no. 1758250). The DNA of the cosmid vector SuperCos1 (Wahl et al. (1987), Proceedings of the
20 National Academy of Sciences, USA 84:2160-2164), obtained from Stratagene (La Jolla, USA, Product Description SuperCos1 Cosmid Vector Kit, Code no. 251301) was cleaved with the restriction enzyme XbaI (Amersham Pharmacia, Freiburg, Germany, Product Description XbaI, Code no. 27-0948-02) and likewise dephosphorylated with shrimp alkaline phosphatase.

25 The cosmid DNA was then cleaved with the restriction enzyme BamHI (Amersham Pharmacia, Freiburg, Germany, Product Description BamHI, Code no. 27-0868-04). The cosmid DNA
30 treated in this way was mixed with the treated ATCC13032-DNA, and the batch was then treated with T4-DNA ligase (Amersham Pharmacia, Freiburg, Germany, Product Description T4-DNA ligase, Code no.27-0870-04). The ligation mixture

was then packed into phages using Gigapack II XL Packing Extracts (Stratagene, La Jolla, USA, Product Description Gigapack II XL Packing Extract, Code no. 200217).

In order to infect the *E. coli* strain NM554 (Raleigh et al. 1988, Nucleic Acid Res. 16:1563-1575) the cells were taken up in 10 mM MgSO₄ and mixed with an aliquot of the phage suspension. Infection and titration of the cosmid library were carried out as described by Sambrook et al. (1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor), the cells being plated out on LB agar (Lennox, 1955, Virology, 1:190) + 100 µg/ml ampicillin. After incubation overnight at 37°C recombinant individual clones were selected.

Example 2

Isolation and sequencing of the gene *lysR1*

The cosmid DNA of an individual colony was isolated with the Qiaprep Spin Miniprep Kit (Product No. 27106, Qiagen, Hilden, Germany) according to the manufacturer's instructions and then partially cleaved with the restriction enzyme Sau3AI (Amersham Pharmacia, Freiburg, Germany, Product Description Sau3AI, Product No. 27-0913-02). The DNA fragments were dephosphorylated with shrimp alkaline phosphatase (Roche Molecular Biochemicals, Mannheim, Germany, Product Description SAP, Product No. 1758250). After gel electrophoresis separation the cosmid fragments were isolated in the size range from 1500 to 2000 bp using the QiaExII Gel Extraction Kit (Product No. 20021, Qiagen, Hilden, Germany).

The DNA of the sequencing vector pZero-1 obtained from Invitrogen (Groningen, Netherlands, Product Description Zero Background Cloning Kit, Product No. K2500-01) was cleaved with the restriction enzyme BamHI (Amersham Pharmacia, Freiburg, Germany, Product Description BamHI,

Product No. 27-0868-04). The ligation of the cosmid fragments into the sequencing vector pZero-1 was carried out as described by Sambrook et al. (1989, Molecular Cloning: A Laboratory Manual, Cold Spring Harbor), the DNA mixture having been incubated overnight with T4 ligase (Pharmacia Biotech, Freiburg, Germany). This ligation mixture was then electroporated into the *E. coli* strain DH5 MCR (Grant, 1990, Proceedings of the National Academy of Sciences, U.S.A., 87:4645-4649) (Tauch et al. 1994, FEMS Microbiol. Letters, 123:343-7) and plated out onto LB-agar (Lennox, 1955, Virology, 1:190) with 50 µg/l zeocin.

The plasmid preparation of the recombinant clone was carried out with the Biorobot 9600 (Product No. 900200, Qiagen, Hilden, Germany). The sequencing was performed according to the dideoxy chain termination method of Sanger et al. (1977, Proceedings of the National Academies of Sciences, U.S.A., 74:5463-5467) as modified by Zimmermann et al. (1990, Nucleic Acids Research, 18:1067). The "RR dRhodamin Terminator Cycle Sequencing Kit" from PE Applied Biosystems (Product No. 403044, Weiterstadt, Germany) was used. The gel electrophoresis separation and analysis of the sequencing reaction were carried out in a "Rotiphoresis NF Acrylamide/Bisacrylamide" Gel (29:1) (Product No. A124.1, Roth, Karlsruhe, Germany) using the "ABI Prism 377" sequencing device from PE Applied Biosystems (Weiterstadt, Germany).

The raw sequence data thus obtained were then processed using the Staden Program Package (1986, Nucleic Acids Research, 14:217-231) Version 97-0. The individual sequences of the pZero1 derivatives were assembled to form a coherent contig. The computer-assisted coding region analysis was prepared using the XNIP program (Staden, 1986, Nucleic Acids Research, 14:217-231). Further analyses were carried out with the "BLAST search programs" (Altschul et al., 1997, Nucleic Acids Research, 25:33893402) against the

non-redundant database of the National Center for
Biotechnology Information (NCBI, Bethesda, MD, USA).

The nucleotide sequence thus obtained is represented in SEQ
ID No. 1. Analysis of the nucleotide sequence revealed an
open reading frame of 912 base pairs, which was termed the
lysR1 gene. The lysR1 gene codes for a polypeptide of 304
amino acids.

Example 3

Production of an integration vector for the integration
mutagenesis of the lysR1 gene

Chromosomal DNA was isolated from the strain ATCC 13032 by
the method of Eikmanns et al. (Microbiology 140: 1817 -
1828 (1994)). On account of the sequence of the lysR1 gene
known from Example 2 for *C. glutamicum*, the following
oligonucleotides were selected for the polymerase chain
reaction:

lysR1intA:

5`TTC CAA TCC CTG CTG TTC AC 3` (SEQ ID NO;4)

lysR1intB:

5`GTG ACC TTT GAA ACC AGC GA 3` (SEQ ID NO:5)

The represented primers were synthesised by MWG Biotech
(Ebersberg, Germany) and the PCR reaction was carried out
according to the standard PCR method of Innis et al. (PCR
protocols. A guide to methods and applications, 1990,
Academic Press) using Pwo polymerase from Boehringer. By
means of the polymerase chain reaction a 383 bp long
internal fragment of the lysR1 gene was isolated, which is
shown in SEQ ID No. 3.

The amplified DNA fragment was ligated into the vector
pCR2.1-TOPO (Mead et al. (1991) Bio/Technology 9:657-663)
using the TOPO TA Cloning Kit from Invitrogen Corporation
(Carlsbad, CA, USA; Cat. No. K4500-01).

The *E. coli* strain TOP10F was then transformed with the ligation batch (Hanahan, In: DNA cloning. A practical approach. Vol. I. IRL-Press, Oxford, Washington DC, USA, 1985). Plasmid-carrying cells were selected by plating out the transformation batch onto LB agar (Sambrook et al., Molecular cloning: a laboratory manual. 2nd Ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989) that had been supplemented with 25 mg/l of kanamycin. Plasmid DNA was isolated from a transformant using the QIAprep Spin Miniprep Kit from Qiagen and was checked by restriction with the restriction enzyme EcoRI followed by agarose gel electrophoresis (0.8%). The plasmid was named pCR2.1lysR1int.

Example 4

Integration mutagenesis of the lysR1 gene in the lysine producer DSM 5715

The vector pCR2.1lysR1int mentioned in Example 3 was electroporated into *Corynebacterium glutamicum* DSM 5715 according to the electroporation method of Tauch et. al. (FEMS Microbiological Letters, 123:343-347 (1994)). The strain DSM 5715 is an AEC-resistant lysine producer. The vector pCR2.1lysR1int cannot replicate independently in DSM 5715 and thus only remains in the cell if it has integrated into the chromosome of DSM 5715. The selection of clones with pCR2.1lysR1int integrated into the chromosome was made by plating out the electroporation batch onto LB agar (Sambrook et al., Molecular cloning: a laboratory manual. 2nd Ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.) that had been supplemented with 15 mg/l of kanamycin.

In order to demonstrate the integration the lysR1int fragment was labelled using the Dig Hybridisation Kit from Boehringer according to the method described in "The DIG System User's Guide for Filter Hybridization" published by

Boehringer Mannheim GmbH (Mannheim, Germany, 1993).

Chromosomal DNA of a potential integrant was isolated according to the method of Eikmanns et al. (Microbiology 140: 1817 - 1828 (1994)) and was in each case cleaved with the restriction enzymes SalI, SacI and HindIII. The resultant fragments were separated by means of agarose gel electrophoresis and hybridised at 68°C using the Dig Hybridisation Kit from Boehringer. The plasmid pCR2.1lysR1int mentioned in Example 3 had inserted itself into the chromosome of DSM 5715 within the chromosomal lysR1 gene. The strain was designated DSM 5715::pCR2.1lysR1int.

Example 5

Production of lysine

The *C. glutamicum* strain DSM 5715::pCR2.1lysR1int obtained in Example 4 was cultivated in a nutrient medium suitable for the production of lysine and the lysine content in the culture supernatant was determined.

For this purpose the strain was first of all incubated for 24 hours at 33°C on an agar plate with the corresponding antibiotic (brain-heart agar with kanamycin (25 mg/l)). Starting from this agar plate culture a preculture was inoculated (10 ml of medium in a 100 ml Erlenmeyer flask). The full medium CgIII was used as medium for the preculture.

Medium Cg III

NaCl	2.5 g/l
Bacto-Peptone	10 g/l
Bacto-Yeast Extract	10 g/l
Glucose (autoclaved separately)	2% (w/v)

The pH value was adjusted to pH 7.4

Kanamycin (25 mg/l) was added to this preculture. The preculture was then incubated for 24 hours at 33°C at 240 rpm on a shaker table. From this preculture a main culture was inoculated so that the initial OD (660 nm) of the main culture was 0.1 OD. The medium MM was used for the main culture.

Medium MM

CSL (Corn Steep Liquor)	5 g/l
MOPS	20 g/l
Glucose (autoclaved separately)	50g/l
Salts:	
(NH ₄) ₂ SO ₄)	25 g/l
KH ₂ PO ₄	0.1 g/l
MgSO ₄ .7H ₂ O	1.0 g/l
CaCl ₂ .2H ₂ O	10 mg/l
FeSO ₄ .7H ₂ O	10 mg/l
MnSO ₄ .H ₂ O	5.0 mg/l
Biotin (sterile filtered)	0.3 mg/l
Thiamine.HCl (sterile filtered)	0.2 mg/l
Leucine (sterile filtered)	0.1 g/l
CaCO ₃	25 g/l

CSL, MOPS and the salt solution are adjusted with ammonia water to pH 7 and autoclaved. The sterile substrate and

vitamin solutions as well as the dry autoclaved CaCO_3 are then added.

Cultivation is carried out in a 10 ml volume in a 100 ml Erlenmeyer flask equipped with baffles. Kanamycin was added (25 mg/l). The cultivation was carried out at 33°C and 80% atmospheric humidity.

After 72 hours the OD was determined at a measurement wavelength of 660 nm with a Biomek 1000 (Beckmann Instruments GmbH, Munich). The amount of lysine formed was determined by ion exchange chromatography and post-column derivatisation with ninhydrin detection using an amino acid analyser from Eppendorf-BioTronik (Hamburg, Germany).

The results of the experiment are shown in Table 1.

Table 1

Strain	OD(660)	Lysine-HCl g/l
DSM 5715	7.5	13.01
DSM 5715:: pCR2.1lysR1int	7.7	15.64

The acronyms and abbreviations used have the following meanings.

KmR: Kanamycin resistance gene

EcoRI: Cleavage site of the restriction enzyme
EcoRI

lysR1int: Internal fragment of the lysR1 gene

ColE1 ori: Replication origin of the plasmid ColE1

Obviously, numerous modifications and variations on the present invention are possible in light of the above teachings. It is therefore to be understood that within the scope of the appended claims, the invention may be practiced otherwise than as specifically described herein.